EFFECT OF REPLACEMENT OF BARLEY GRAINS AND SOYBEAN MEAL BY DISTILLER’S DRIED GRAINS WITH SOLUBLES WITH OR WITHOUT SUPPLEMENTED SEAWEED IN GROWING RABBIT RATIONS ON: 2. Calcium and phosphorus intake and absorption and some blood constituents of growing rabbits.

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ABSTRACT

Twenty seven of weaning New Zealand White (NZW) rabbits of seven weeks of age was randomly distributed into nine groups of equal number (three rabbits in each) and similar average live body weight (813g ± 0.01). Each group at 7, 9, 11 and 13 weeks of age was tended to determine Ca, and P concentration in blood plasma. At 13 weeks old, blood samples were collected from the ear vein of the three rabbits in each group after overnight fasting during the last day of all growth trials. A representative part (5cm) from the small intestine was dissected immediately after slaughtering of rabbits to determine Ca and P absorption rates.

The experimental groups were fed randomly on one of the nine formulated experimental rations used. The 1\textsuperscript{st} ration (R1) was used as a control, which contained 10 % yellow corn + 10 % barley + 13.7 % soybean meal (SBM) +20 % wheat bran +40 % clover hay + 3 % molasses + 1 % dicalcium phosphate + 1.2 % limestone + 0.5 % sodium chloride + 0.4 % premix + 0.2 % methionine and substituting the equal parts of barley and SBM by 10 % and 20 % distiller's dried grains with solubles (DDGS) for ration 2 (R2) and ration 3 (R3), respectively. The supplemented seaweed (SW) for these rations was at two levels. The first level was 0.5 % seaweed of the total mixed ration for ration 4 (R4), ration 5 (R5) and ration 6 (R6). The second level was 1.0 % seaweed of the total
mixed ration for ration 7 (R7), ration 8 (R8) and ration 9 (R9). All rations were in pelleted form and nearly isonitrogenous and isocaloric.

The results of the present study revealed that Ca intake was increased (P < 0.05) with feeding DDGS on R8 (1.92 g/h/d) than the other rations, while feeding on R2, R7 and R9 decreased (P < 0.05) Ca intake (1.67, 1.76 and 1.07 g/h/d, respectively) at 12 to 13 weeks of age. Phosphorous intake was increased (P < 0.05) with feeding on R3, R5, R6 and R8 (0.96, 0.94, 0.95 and 0.96 g/h/d, respectively). The mean values from 7 to 13 weeks old ranged from 0.61 to 0.71 g/h/d of the experimental rations. There was no significant effect with feeding on DDGS with or without SW on Ca and P absorption and concentration in blood plasma.

Urea-N concentration in blood plasma was increased (P < 0.05) with feeding on R2, R3 and R7 (45.95, 45.37 and 46.53 mg/100ml, respectively) than the other ration groups.

The feeding R9 was higher (P<0.05) in cholesterol concentration (224.68 mg/100ml) than the other rations, while feeding on R3, R5 and R6 were higher (P < 0.05) (202.22, 213.84 and 215.39 mg/100ml, respectively) than the others. Triglycerides concentration was higher (P < 0.05) with feeding R3, R5, R6, R8 and R9 (107.26, 122.02, 115.62, 119.56 and 133.58 mg/100ml, respectively) than the other rations, and total lipids was also higher with the same rations (430.0, 225.0, 267.5, 232.5 and 282.5 mg/100ml, respectively). Low density lipoproteins (LDL) concentration was higher (P < 0.05) with feeding on R9 (164.23 mg/100ml) than the other rations.

It could be concluded that feeding on DDGS without SW supplementation could be increased urea-N in blood plasma, triglycerides, cholesterol and total lipids in blood plasma of growing rabbits.

**Keywords:** Calcium, phosphorus, blood, growing rabbits, DDGS, SBM, seaweed.

**INTRODUCTION**

One of the more commonly held thoughts on calcium metabolism in rabbits is that the rabbit's gastrointestinal system is designed to absorb all the dietary calcium that is presented to it (Rosenthal, 2006). In other words, the more calcium in the food, the more calcium enters the gastrointestinal tract. Eventually, the theory goes, excess calcium in the body can not stay in the blood stream and is excreted though the urinary tract. This theory postulates that there is no delicate balancing act among calcium, phosphorous, vitamin D and parathyroid hormone
(PTH) found in all other mammals. Without this ballet act among these four components, there is no gateway to prevent an overabundance of calcium absorption through the gastrointestinal tract. It is also thought that great majority of calcium absorbed by the gastrointestinal tract leaves the body through the urinary tract. This differs from almost all other mammals as studies and show in most animals other than rabbit’s in which calcium is excreted, harmlessly through fecal material.

When calcium is measured in dietary experiments, it is important to look at the relationship of calcium to phosphorous. A limiting factor in the use of digestible P in diet formulations is accuracy of bioavailability estimates for feedstuffs (Robbins et al., 2000). Therefore, in diets with higher energy contents, it may be necessary to raise the concentration of P. Ultimately, P requirements for market animals should probably be based on the amount of P required per pound of lean tissue growth.

Monogastric animals do not secrete phytase in sufficient quantities to breakdown the phytate molecule; hence most of phytate P is not available for absorption (Soares and Jr., 1995). Therefore, large amounts of a highly available inorganic P source must be added to meet the P requirement. While most phytate P is not available for absorption, much of this phytate P is mineralized to inorganic P in large intestine, and is excreted in manure as inorganic P and increase soluble P loss potential when applied to soils (Soares and Jr., 1995).

Rabbit rearing is an established micro livestock industry in many countries where rabbit are domesticated for meat. Broiler breeds of rabbits have also been introduced in India to explore its avenue as an alternative source of animal protein. Rabbit meat has high biological value (21%) and low in fat and cholesterol (Sinha et al., 2008).

There is a tradition of rabbit production in the five Mediterranean countries of Africa. Per caput production varies from Egypt’s 0.27 Kg to Morocco’s nearly 0.78 Kg (Lebas et al., 1997).

Rabbits grow rapidly and its rate is comparable to that of broiler chicken (Sinha et al., 2008). Proper nutrition is one of the important aspects of broiler rabbit production. The protein and energy content of the diet play a vital role in rabbit nutrition. Biochemical characterization of rabbit will help in better understanding of rabbit in relation to growth, meat, and fur quality.

By-product feed can serve as a source of nutrients in animal diets. Often, byproduct feed such as Distillers grains are included in the diet of they are readily available and economically justified, especially, when there is a shortage or increase in prices of conventional feed sources. Also, due the processing methods employed, nutrients in the by products become more biologically available and
can potentially reduce nutrient excretion if the by product nutrient can be balanced in the diet (Spiehs et al., 2002).

Distiller's dried grains with soluble (DDGS) which contains from 0.62 to 0.87 % P, have a higher concentration of available P than corn, other cereal grains and cereal co-products, averaging 77 % for DDGS compared to a range of 12 to 30 % for corn. Studies by Spiehs et al. (2002) showed that when formulating diets on a total P basis, the percentage of P retained tends to increase when 10 and 20 % DDGS are added to growing diets compared to a control corn-soybean meal diets (63.9 %, 66.3 % and 59.1 %, respectively).

Although the DDGS contains a significant amount of crude fiber (8 – 10 %), it also contains 10 – 12 % fat (Whitney et al., 1999). Fats as energy carriers and sources of essential unsaturated fatty acids have attracted the attention of nutritionists in recent years. The activity of these acids in the animal body is reflected mainly in the activity of eicosanoids (known as tissue hormones). Owing to the mechanism of their action, they can be treated as the most peripheral first messengers, which strengthen or weaken the regulatory activity of hormones and neuromediators at cellular level (Corl et al., 2003).

The chemical composition of ordinary seaweed as from Ascophyllum nodosum, immediately characterizes the material as of low-energy content. According to the analytical data the value of seaweed meal must primarily be sought in its content of vitamins and mineral among which β-carotene, tocopherols, some B vitamins, iodine, zinc and potassium are the more important components (Scott, 1990).

Mineral and vitamins are also of great importance in rabbit diets. It should be noted though that the rabbit cannot regulate its calcium absorption. Imbalances in calcium and phosphorous nutrition (too much P per unit Ca) can lead to dental problems and urinary tract obstructions (Urolithiasis).

Therefore, the objective of this study was to evaluate the effect of partially or totally substituting of barley and partially of soybean meal by DDGS with or without seaweed supplementation on calcium and phosphorus absorption and some blood constituents of growing NZW rabbits and their effects on growth rate through Ca and P uptake and blood parameters.

**MATERIALS AND METHODS**

The experimental field of the present study was carried out at the Experimental Station of the Poultry Production Department, while, the chemical analysis was run at the Laboratory of the Animal Production Department, Faculty of Agriculture, Mansoura University, Mansoura, Egypt.
**Experimental animals:**

Twenty seven of weaning New Zealand White (NZW) rabbits of seven weeks old was randomly distributed into nine groups of equal number (three rabbits in each) and similar average live body weight (813g ± 0.01). All rabbits of each group at 7, 9, 11 and 13 weeks of age were tended to determined Ca, and P concentration in blood plasma. At 13 weeks old, blood samples were collected from the ear vein of the rabbits in each group after overnight fasting during the last day of all growth trials.

**Treatments and experimental design:**

The experimental groups were fed randomly on one of the nine formulated experimental rations used. The experimental rations were designed to gradually substitute barley grains and soybean meal (equal parts) by distiller dried grains with solubles (DDGS) at the rate of 10 and 20 % of the total mixed rations. Ration 1 (R1) contains 10 % barley and 13.7 % SBM as control diets, and the substituting the barley and SBM by 10 % and 20 % DDGS for ration 2 (R2) and ration 3 (R3), respectively. The supplemented seaweed for these rations was at two levels of the wheat bran. The first level was by 0.5 % seaweed (SW) of the total mixed ration for ration 4 (R4), ration 5 (R5) and ration 6 (R6). The second level was by 1.0 % seaweed of the total mixed ration for ration 7 (R7), ration 8 (R8) and ration 9 (R9). All rations were in pelleted form and nearly isonitrogenous and isocaloric content.

**Blood constituents’ measurements.**

Blood samples were collected from the ear vein of the three rabbits in each group at 7, 9, 11 and 13 weeks old to determine Ca, according to Moorhead and Briggs (1974) and P, according to Freidman *et al.* (1980). At 13 weeks old, blood samples were collected from the ear vein of rabbits in each group after overnight fasting during the last day of all growth trial. Blood plasma was separated after centrifugation at 4000 r.p.m. for 20 minutes, and then stored at −20°C until analysis for the different blood parameters. Plasma was used for determination of total proteins, (Doumas *et al.*, 1981); albumin, (Hill and Wells, 1983); globulin, which calculated by difference between the total proteins and albumin concentrations); AST and ALT, (Reitman and Frankel, 1957); alkaline phosphates, (Roy, 1970); urea-N, (Freidman *et al.*, 1980); uric acid, (Fossati *et al.*, 1980); creatinine (Fabiny and Ertingshausen, 1971); cholesterol(Allain *et al.*, 1974); triglycerides (Fossati and Prencipe, 1982); total lipids (Zollner and Kirsch, 1962); HDL, (Lopes-Virella *et al.*, 1977); LDL, (Friedewald *et al.*, 1972), were carried out using photometric methods and diagnostic kits (Vitro Scient, Egypt), and pH determined by battery operated pH meter.
Absorption of Ca and P (in vitro) from intestine:
A representative part (5 cm) from the small intestine was dissected immediately after slaughtering of rabbits. The dissected tissues were immersed in 20 ml of CaHPO$_4$ solution in Petri dishes for one hour. After that the solution was collected in clean tubes and then used to determine Ca and P absorption rates. Absorption rates were calculated by subtracting the detected amounts in the collected samples from that in the immersed solutions. The methods of Ca and P determination were the same as those used for blood plasma samples by using available commercial kits.

Statistical analysis:
The statistical analysis was carried out using the General Linear Model Program (GLM) of SAS (2000). The obtained data were analyzed using factorial analysis (3x3) of variance according to the following model:

\[ Y_{ijk} = \mu + T_i + L_j + TL_{ij} + e_{ijk} \]

Where; \( Y_{ijk} \) = Observation of the tested factor, \( \mu \) = Overall mean, \( T_i \) = The effect of treatment (DDGS), \( i = 0, 10 \text{ and } 20 \% \), \( L_j \) = The effect of levels (seaweed), \( j = 0, 0.5 \text{ and } 1 \% \), \( TL_{ij} \) = The interaction between treatment and level effect and \( e_{ijk} \) = The random error associated with the individual ijk.

Differences among means were subjected to Duncan’s Multiple Range Test (Duncan, 1955).

RESULTS AND DISCUSSION

Effect of feeding with DDGS or SW supplementation and their interaction on Ca and P intake:
Table 1 shows that there was no significant effect of supplemented SW on Ca and P intake. The Ca and P intake increased with increasing the age from 7 to 13 weeks of age. These values of Ca intake were (0.87, 0.87 and 0.90 g/h/d) and P intake were (0.44, 0.43 and 0.45 g/h/d) at 7 to 8 weeks of age, while the values of Ca intake were (1.79, 1.84 and 1.8 g/h/d) and for P intake were (0.09, 0.92 and 0.90 g/h/d) at 12 to 13 weeks with supplemented 0.0, 0.5 and 1.0 % SW, respectively.

Data in Table 1 show that feeding on DDGS, data show that the Ca intake was increased (P < 0.05) (1.78 g/h/d) with feeding on 20 % DDGS than the control (1.57 g/h/d), but there was no significant effect with feeding on 10 % DDGS (1.68 g/h/d) and control or with 20 % DDGS at 11 to 12 week old. The mean values of
EFFECT OF DDGS OR SWDEITS ON BLOOD CONSTITUENTS OF RABBITS

Table 1. Main effects of SW supplementation or feeding DDGS on Ca and P intake (g/h/d) of growing rabbits.

<table>
<thead>
<tr>
<th>Items</th>
<th>SW(%)</th>
<th>DDGS(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0.0±SEM</td>
<td>P</td>
</tr>
<tr>
<td>Wk (7 – 8)</td>
<td>0.87</td>
<td>0.87</td>
</tr>
<tr>
<td>Wk (8 – 9)</td>
<td>1.01</td>
<td>0.99</td>
</tr>
<tr>
<td>Wk (9 – 10)</td>
<td>1.11</td>
<td>1.12</td>
</tr>
<tr>
<td>Wk (10 – 11)</td>
<td>1.51</td>
<td>1.46</td>
</tr>
<tr>
<td>Wk (11 – 12)</td>
<td>1.69</td>
<td>1.66</td>
</tr>
<tr>
<td>Wk (12 – 13)</td>
<td>1.79</td>
<td>1.84</td>
</tr>
<tr>
<td>Mean</td>
<td>1.33</td>
<td>1.32</td>
</tr>
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</table>

<table>
<thead>
<tr>
<th>Items</th>
<th>SW(%)</th>
<th>DDGS(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0.0±SEM</td>
<td>P</td>
</tr>
<tr>
<td>Wk (7 – 8)</td>
<td>0.44</td>
<td>0.43</td>
</tr>
<tr>
<td>Wk (8 – 9)</td>
<td>0.50</td>
<td>0.49</td>
</tr>
<tr>
<td>Wk (9 – 10)</td>
<td>0.56</td>
<td>0.56</td>
</tr>
<tr>
<td>Wk (10 – 11)</td>
<td>0.75</td>
<td>0.73</td>
</tr>
<tr>
<td>Wk (11 – 12)</td>
<td>0.85</td>
<td>0.83</td>
</tr>
<tr>
<td>Wk (12 – 13)</td>
<td>0.90</td>
<td>0.92</td>
</tr>
<tr>
<td>Mean</td>
<td>0.67</td>
<td>0.66</td>
</tr>
</tbody>
</table>

Ca intake were (1.31, 1.34 and 1.34 g/h/d) with feeding on 0.0 %, 10 % and 20 % DDGS, respectively. Phosphorous intake was increased (P < 0.05) with feeding on 10 or 20 % DDGS. The values for P intake were (0.42, 0.44 and 0.46 g/h/d) at 7 to 8 week and (0.87, 0.91 and 0.93 g/h/d) at 12 to 13 week, while the mean values were (0.63, 0.67 and 0.70 g/h/d) for 7 to 13 weeks with feeding on 0.0 %, 10 % and 20 % DDGS, respectively.

Obtained results as shown in Table 2, present the interaction effect of feeding on DDGS with or without SW, show that the Ca intake was increased (P < 0.05) with feeding on R8 (1.92 g/h/d) than the other rations, while feeding on R2, R7 and R9 were decreased (P< 0.05) Ca intake (1.67, 1.76 and 1.07 g/h/d, respectively) at 12 to 13 weeks old. The mean values from 7 to 13 weeks of age were ranged from 1.27 to 1.37 g/h/d. Phosphorous intake increased (P < 0.05) with feeding on R3, R5, R6 and R8 (0.96, 0.94, 0.95 and 0.96 g/h/d, respectively). The mean values of 7 to 13 weeks old ranged from (0.61 to 0.71 g/h/d).
## Table 2. Interaction effects between feeding on DDGS and with or without SW on Ca and P intake (g/h/d) of growing rabbits.

<table>
<thead>
<tr>
<th>Items</th>
<th>R1</th>
<th>R2</th>
<th>R3</th>
<th>R4</th>
<th>R5</th>
<th>R6</th>
<th>R7</th>
<th>R8</th>
<th>R9</th>
<th>± SEM</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>DDGS(%)</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td></td>
<td></td>
</tr>
<tr>
<td>SW(%)</td>
<td>0.0</td>
<td>0.5</td>
<td>1.0</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Ca intake (g/h/d):</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wk (7 – 8)</td>
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<td>0.89</td>
<td>0.86</td>
<td>0.84</td>
<td>0.88</td>
<td>0.88</td>
<td>0.90</td>
<td>0.89</td>
<td>0.92</td>
<td>0.023</td>
<td>0.659</td>
</tr>
<tr>
<td>Wk (8 – 9)</td>
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<td>1.02</td>
<td>1.00</td>
<td>0.98</td>
<td>0.99</td>
<td>1.01</td>
<td>1.03</td>
<td>1.00</td>
<td>1.00</td>
<td>0.021</td>
<td>0.471</td>
</tr>
<tr>
<td>Wk (9 – 10)</td>
<td>1.12</td>
<td>1.11</td>
<td>1.11</td>
<td>1.11</td>
<td>1.13</td>
<td>1.12</td>
<td>1.13</td>
<td>1.14</td>
<td>1.12</td>
<td>0.020</td>
<td>0.831</td>
</tr>
<tr>
<td>Wk (10 – 11)</td>
<td>1.55</td>
<td>1.52</td>
<td>1.46</td>
<td>1.40</td>
<td>1.49</td>
<td>1.49</td>
<td>1.52</td>
<td>1.52</td>
<td>1.51</td>
<td>0.041</td>
<td>0.313</td>
</tr>
<tr>
<td>Wk (11 – 12)</td>
<td>1.60</td>
<td>1.61</td>
<td>1.86</td>
<td>1.49</td>
<td>1.68</td>
<td>1.81</td>
<td>1.63</td>
<td>1.75</td>
<td>1.68</td>
<td>0.087</td>
<td>0.334</td>
</tr>
<tr>
<td>Wk (12 – 13)</td>
<td>1.85</td>
<td>1.67</td>
<td>1.86</td>
<td>1.88</td>
<td>1.83</td>
<td>1.76</td>
<td>1.92</td>
<td>1.70</td>
<td>0.059</td>
<td>0.027</td>
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<tr>
<td><strong>Mean</strong></td>
<td>1.33</td>
<td>1.30</td>
<td>1.36</td>
<td>1.27</td>
<td>1.34</td>
<td>1.35</td>
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<td>1.37</td>
<td>1.32</td>
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<td>0.083</td>
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<tr>
<td><strong>P intake (g/h/d):</strong></td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
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</tr>
<tr>
<td>Wk (7 – 8)</td>
<td>0.42</td>
<td>0.45</td>
<td>0.45</td>
<td>0.40</td>
<td>0.44</td>
<td>0.46</td>
<td>0.43</td>
<td>0.45</td>
<td>0.48</td>
<td>0.011</td>
<td>0.668</td>
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<tr>
<td>Wk (8 – 9)</td>
<td>0.48</td>
<td>0.51</td>
<td>0.52</td>
<td>0.47</td>
<td>0.49</td>
<td>0.52</td>
<td>0.50</td>
<td>0.50</td>
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<td>0.010</td>
<td>0.515</td>
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<td>Wk (9 – 10)</td>
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<td>0.55</td>
<td>0.58</td>
<td>0.53</td>
<td>0.57</td>
<td>0.58</td>
<td>0.54</td>
<td>0.57</td>
<td>0.58</td>
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<td>0.852</td>
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<td>Wk (10 – 11)</td>
<td>0.74</td>
<td>0.76</td>
<td>0.76</td>
<td>0.67</td>
<td>0.75</td>
<td>0.77</td>
<td>0.73</td>
<td>0.76</td>
<td>0.78</td>
<td>0.021</td>
<td>0.361</td>
</tr>
<tr>
<td>Wk (11 – 12)</td>
<td>0.77</td>
<td>0.80</td>
<td>0.97</td>
<td>0.71</td>
<td>0.84</td>
<td>0.94</td>
<td>0.78</td>
<td>0.87</td>
<td>0.87</td>
<td>0.043</td>
<td>0.320</td>
</tr>
<tr>
<td>Wk (12 – 13)</td>
<td>0.89</td>
<td>0.84</td>
<td>0.96</td>
<td>0.87</td>
<td>0.94</td>
<td>0.95</td>
<td>0.85</td>
<td>0.96</td>
<td>0.88</td>
<td>0.030</td>
<td>0.028</td>
</tr>
<tr>
<td><strong>Mean</strong></td>
<td>0.64</td>
<td>0.65</td>
<td>0.71</td>
<td>0.61</td>
<td>0.67</td>
<td>0.70</td>
<td>0.64</td>
<td>0.69</td>
<td>0.69</td>
<td>0.011</td>
<td>0.080</td>
</tr>
</tbody>
</table>

SEM = Standard error of means,  
*P* = Probability.  
Wk = Week

The present results were in agreement with those of authors who reported that growing rabbits should be fed to appetite until 10 weeks of age and then at around 112 g DM/day (Santoma et al., 1989). The Ca and P requirements of growing rabbits are 1.15 and 0.6 %, respectively (Moughan et al., 1988). Where, the Ca and P intake of growing rabbits were (1.30 and 0.67 g/h/d, respectively).

### Effect of SW supplementation or feeding DDGS and their interaction on Ca (mol/hr) and P (mol/hr) absorption:

Table (3) shows the absorption rate of Ca and P at 13 weeks of age. The results showed that there were no significant effect of with SW or

## Table 3. Main effects of SW supplementation or feeding DDGS on Ca (mol/hr) and P (mol/hr) absorption at week 13 of age.

<table>
<thead>
<tr>
<th>Items</th>
<th>SW(%)</th>
<th>DDGS(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0.0</td>
<td>0.5</td>
</tr>
<tr>
<td>Ca (mol/hr)</td>
<td>2.71</td>
<td>2.65</td>
</tr>
<tr>
<td>P (mol/hr)</td>
<td>1.12</td>
<td>1.15</td>
</tr>
</tbody>
</table>

SEM = Standard error of means,  
*P* = Probability.
feeding on DDGS on Ca and P absorption. The mean values for Ca absorption ranged from 2.65 to 2.76 (mol/hr) and for P absorption ranged from 1.10 to 1.16 (mol/hr).

There was no significant interaction effect with feeding on DDGS with or without SW on Ca and P absorption. As shown in Table (4), the mean values for Ca absorption ranged from 2.57 to 2.94 (mol/hr) and for P absorption were ranged from 1.06 to 1.25 (mol/hr) with feeding on the experimental rations.

Carbohydrates increased intestinal Ca\(^{2+}\) absorption as reported by (Buzinaro et al., 2006). Data regarding protein intake and intestinal Ca\(^{2+}\) absorption indicate that dietary protein does not alter the intestinal Ca\(^{2+}\) absorption and hence intestinal calcium absorption does not explain hypercalcuria induced by high protein intake (Kerstetter et al., 2003). However, the data related to the effect of dietary lipids on intestinal Ca\(^{2+}\) absorption are not clear (Buzinaro et al., 2006). In addition, a high phosphorus intake has been showed to cause hypocalcemia, hyperphosphatemia, secondary hyperparathyroidism with enhanced bone resorption and bone loss in several animal models (Calvo and Park, 1996).

Table 4. Interaction effects between feeding on DDGS with or without SW on Ca (mol/hr) and P (mol/hr) absorption at 13 weeks of age.

<table>
<thead>
<tr>
<th>Items</th>
<th>R1</th>
<th>R2</th>
<th>R3</th>
<th>R4</th>
<th>R5</th>
<th>R6</th>
<th>R7</th>
<th>R8</th>
<th>R9</th>
<th>± SEM</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>DDGS(%)</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td>0.0</td>
<td>10.0</td>
<td>20.0</td>
<td>0.216</td>
<td>0.803</td>
</tr>
<tr>
<td>SW(%)</td>
<td>0.0</td>
<td>0.5</td>
<td>1.0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ca (mol/hr)</td>
<td>2.75</td>
<td>2.67</td>
<td>2.71</td>
<td>2.65</td>
<td>2.57</td>
<td>2.74</td>
<td>2.75</td>
<td>2.94</td>
<td>2.60</td>
<td>0.216</td>
<td>0.803</td>
</tr>
<tr>
<td>P (mol/hr)</td>
<td>1.06</td>
<td>1.14</td>
<td>1.16</td>
<td>1.25</td>
<td>1.06</td>
<td>1.15</td>
<td>1.16</td>
<td>1.12</td>
<td>1.09</td>
<td>0.103</td>
<td>0.718</td>
</tr>
</tbody>
</table>

SEM = Standard error of means.  
P = Probability.

Phosphorus balance involves the absorption of dietary phosphorus in the intestine, its distribution in body fluids and tissues, especially bone and its excretion was largely by the kidney (Tani et al., 2007). Intestinal net phosphorus absorption was significantly increased by the high phosphorus diets, dependent on the amount of dietary phosphorus.

Effect of SW supplementation or feeding DDGS and their interaction on Ca and P concentration (mg / 100ml) in blood plasma:

As shown in Table 5, calcium concentration was increased (P < 0.05) with supplemented 1 % SW (9.42 mg/100ml) than supplemented 0.0 or 0.5 % SW (5.16 and 7.86 mg/100ml, respectively), while Ca concentration was higher (P < 0.05) when supplemented 0.5 % SW than the control at 11 weeks of age.
Table 5. Main effects SW supplementation or feeding DDGS on Ca and P concentration (mg /100ml) in blood plasma from 7 to 13 weeks of age.

<table>
<thead>
<tr>
<th>Items</th>
<th>SW(%)</th>
<th>DDGS(%)</th>
<th>0.0 ± SEM</th>
<th>0.5</th>
<th>1.0</th>
<th>0.0 ± SEM</th>
<th>10.0</th>
<th>20.0</th>
<th>± SEM</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ca</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>WK (7)</td>
<td>3.90</td>
<td>3.99</td>
<td>3.86</td>
<td>0.191</td>
<td>0.885</td>
<td>3.92</td>
<td>4.02</td>
<td>3.81</td>
<td>0.191</td>
<td>0.736</td>
</tr>
<tr>
<td>WK (9)</td>
<td>4.47</td>
<td>4.39</td>
<td>3.86</td>
<td>0.262</td>
<td>0.229</td>
<td>4.05</td>
<td>4.34</td>
<td>4.33</td>
<td>0.262</td>
<td>0.676</td>
</tr>
<tr>
<td>WK (11)</td>
<td>5.16</td>
<td>7.86</td>
<td>9.42</td>
<td>0.372</td>
<td>6.19</td>
<td>7.20</td>
<td>9.04</td>
<td>0.372</td>
<td>0.001</td>
<td></td>
</tr>
<tr>
<td>WK (13)</td>
<td>9.66</td>
<td>9.19</td>
<td>9.35</td>
<td>0.576</td>
<td>0.841</td>
<td>9.46</td>
<td>9.46</td>
<td>9.33</td>
<td>0.576</td>
<td>0.987</td>
</tr>
<tr>
<td>Mean</td>
<td>5.80</td>
<td>6.36</td>
<td>6.62</td>
<td>0.184</td>
<td>0.016</td>
<td>5.91</td>
<td>6.24</td>
<td>6.63</td>
<td>0.184</td>
<td>0.042</td>
</tr>
<tr>
<td>P</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>WK (7)</td>
<td>1.52</td>
<td>1.29</td>
<td>1.45</td>
<td>0.509</td>
<td>0.950</td>
<td>1.43</td>
<td>1.37</td>
<td>1.46</td>
<td>0.509</td>
<td>0.992</td>
</tr>
<tr>
<td>WK (9)</td>
<td>2.12</td>
<td>2.36</td>
<td>4.97</td>
<td>0.498</td>
<td>0.001</td>
<td>3.04</td>
<td>2.82</td>
<td>3.59</td>
<td>0.498</td>
<td>0.543</td>
</tr>
<tr>
<td>WK (11)</td>
<td>5.51</td>
<td>3.71</td>
<td>4.37</td>
<td>0.507</td>
<td>0.064</td>
<td>4.32</td>
<td>4.31</td>
<td>4.95</td>
<td>0.507</td>
<td>0.600</td>
</tr>
<tr>
<td>WK (13)</td>
<td>6.52</td>
<td>6.87</td>
<td>7.08</td>
<td>0.424</td>
<td>0.647</td>
<td>6.85</td>
<td>7.48</td>
<td>6.13</td>
<td>0.424</td>
<td>0.109</td>
</tr>
<tr>
<td>Mean</td>
<td>3.92</td>
<td>3.56</td>
<td>4.47</td>
<td>0.227</td>
<td>0.034</td>
<td>3.91</td>
<td>4.00</td>
<td>4.03</td>
<td>0.227</td>
<td>0.928</td>
</tr>
</tbody>
</table>

a, b, c : Means within the same raw with different superscripts are significantly different (P<0.05). SEM = Standard error of means, P = Probability, Wk =Week

The mean values, showed that supplemented 1 % was higher (P<0.05) with supplemented 0.5 % or 1 % SW (6.36 and 6.62 mg/100ml, respectively) than the control group (5.80 mg/100ml).

Phosphorous concentration increased (P<0.05) at 9 week with supplemented 1 % SW (4.97 mg/100 ml) than the control (2.12 mg/100 ml) or supplemented 0.5 % SW (2.36 mg/100ml). The mean values showed that the P concentration was higher (P<0.05) with supplemented 1 % SW (4.47 mg/100ml) than with feeding on 0.5 % SW (3.56 mg/100ml), while there was no significant effect between control (3.92 mg/100ml) and supplemented 0.5 % or 1 % SW.

Feeding on 20 % DDGS increased (P< 0.05) Ca concentration (9.04 mg/100ml) than the control (9.16 mg/100ml) or feeding on 10 % DDGS (7.20 mg/100ml). There was no significant effect on P concentration with feeding on the control or 10 % or 20 % DDGS (3.91, 4.0 and 4.03 mg/100ml, respectively).

As shown in Table 6, there was no significant interaction effect on the mean values for Ca and P concentration in blood plasma. Feeding on R6, R7, R8 and R9 increased Ca concentration (6.87, 6.39, 6.50 and 6.98 mg/100ml, respectively) than the other rations, while feeding on R1, R8 and R9 increased P concentration (4.29, 4.26 and 5.16 mg/100ml, respectively) than the other rations.
Table 6. Interaction effects between feeding on DDGS and with or without SW on Ca and P concentration (mg / 100ml) in blood plasma from 7 to 13 weeks of age.

<table>
<thead>
<tr>
<th>Items</th>
<th>R1</th>
<th>R2</th>
<th>R3</th>
<th>R4</th>
<th>R5</th>
<th>R6</th>
<th>R7</th>
<th>R8</th>
<th>R9</th>
<th>± SEM</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>DDGS(%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.0</td>
<td>3.94</td>
<td>4.24</td>
<td>3.53</td>
<td>4.01</td>
<td>3.99</td>
<td>3.97</td>
<td>3.82</td>
<td>3.83</td>
<td>3.92</td>
<td>0.331</td>
<td>0.787</td>
</tr>
<tr>
<td>10.0</td>
<td>3.33</td>
<td>5.48</td>
<td>4.58</td>
<td>4.69</td>
<td>4.19</td>
<td>4.29</td>
<td>4.12</td>
<td>3.34</td>
<td>4.12</td>
<td>0.454</td>
<td>0.035</td>
</tr>
<tr>
<td>20.0</td>
<td>3.42</td>
<td>5.63</td>
<td>6.42</td>
<td>6.60</td>
<td>6.51</td>
<td>10.46</td>
<td>8.57</td>
<td>9.47</td>
<td>10.23</td>
<td>0.644</td>
<td>0.091</td>
</tr>
<tr>
<td>SW(%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.0</td>
<td>9.93</td>
<td>9.46</td>
<td>9.60</td>
<td>9.41</td>
<td>9.39</td>
<td>8.76</td>
<td>9.05</td>
<td>9.37</td>
<td>9.64</td>
<td>0.997</td>
<td>0.968</td>
</tr>
<tr>
<td>0.5</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>1.0</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Calcium and phosphorous concentrations in blood were within the normal range as found by (Nichols, 2003). Because rabbits can absorb Ca without the facilitation of vitamin D, a mechanism is needed to regulate serum Ca levels. Parathyroid hormone and calcitonin are thought prevent serum Ca level from becoming to dangerously high due to dietary influence (Cheeke, 1994). It is also thought that great majority of calcium absorbed by the gastrointestinal tract leaves the body through the urinary tract (Rosenthal, 2006).

Phosphorous (P) is an essential element of both plant and animals (Schindler, 1977). Therefore, in diets with higher energy contents, it may be necessary to raise the concentration of P. Ultimately, P requirements for market animals should probably be based on the amount of P required per pound of lean tissue growth. A limiting factor in the use of digestible P in diet formulations is accuracy of P bioavailability estimates for feedstuffs (Robbins et al., 2000).

Effect of SW supplementation or feeding DDGS and their interaction on some blood plasma parameters:

As shown in Table 7, there was no significant effect on total protein concentration with supplemented SW or feeding DDGS. Supplemented SW at 1.0 % increased (P<0.05) albumin concentration (4.47 g/100ml) than the control.
EFFECT OF DDGS OR SWDEITS ON BLOOD CONSTITUENTS OF RABBITS

(3.84 g/100ml), while there was no significant effect between supplemented 0.5 % SW (4.0 g/100ml) and the control or supplemented 1 % SW. The AST activity was higher (P<0.05) with supplemented 0.5 % SW (51.11 U/ml) than the control (34.93 U/ml) or 1 % SW (37.24 U/ml), while feeding on 10 % or 20 % DDGS caused increasing (P < 0.05) AST activity (46.04 and 46.84 U/ml, respectively) than the control (30.40 U/ml). Alkaline phosphates (ALP) was higher (P < 0.05) with supplemented 1 % SW (75.26 U/100ml) than the control (53.48 U/100ml), while there was no significant effect between supplemented 0.5 % SW (65.12 U/100ml) and control or supplemented 1 % SW. Creatinine concentration increased (P < 0.05) with supplemented 0.5 % and 1 % SW (1.09 and 1.20 mg/100ml, respectively) than the control (0.95 mg/100ml) or with feeding on 20 % DDGS (1.14 mg/100ml) than feeding on 0.0 % and 10 % DDGS (1.05 mg/100ml for each one).

The interaction results in Table 8 show that urea-N concentration was increased (P<0.05) with feeding on R2, R3 and R7 (45.95, 45.37 and 46.53 mg/100ml, respectively) than the other rations. The present data were within the range as found by Igwebuike et al. (2008) and EL-Banna et al. (2005) except for globulin concentration with feeding on R1, R3, R5, R6, R7 and R8 which were lower than the normal range (2.15 – 3.15 g/100m). EL-Banna et al. (2005) found that the total protein level in plasma of rabbits consumed seaweed was slightly lower than that in the control group and in the meantime higher levels in AST and ALT enzymes in the treated groups. This could be due to increase of protein utilization and amino acids transamination in the treated groups.

Zinc and copper accumulate mostly in the liver tissue (Saito, 1996). However, there are other studies, which suggest that zinc and copper loading may lead to intoxication and increase liver enzymes levels (Saito, 1996 and Levengood et al., 2000) which may support the increase of AST and ALT. Serum alkaline phosphates (ALP) levels may increase in congestive heart failure as a result of injury to the liver (Harper et al., 1977). Changes in the dietary calcium level have been shown to have a profound influence on bone composition, parathyroid size and bone and plasma alkaline phosphates. Increasing enzyme levels in blood plasma resulting from a dietary calcium deficiency (Hurwitz and Griminger, 1973).

Feeding on R9 was higher (P<0.05) in cholesterol concentration (224.68 mg/100ml) than the other rations, while feeding on R3, R5 and R6 were higher (P < 0.05) (202.22, 213.84 and 215.39 mg/100ml, respectively) than the others. Triglycerides concentration was higher (P < 0.05) with feeding on R3, R5, R6, R8 and R9 (107.26, 122.02, 115.62, 119.56 and 133.58 mg/100ml, respectively) than the other rations, and total lipids was also higher with the same relations (430.0,
EFFECT OF DDGS OR SWDEITS ON BLOOD CONSTITUENTS OF RABBITS

225.0, 267.5, 232.5 and 282.5 mg/100ml, respectively). Low density lipoproteins (LDL) concentration was higher (P < 0.05) with feeding on R9 (164.23 mg/100ml) than the other rations, while feeding on R5, R6 and R7 were higher (P < 0.05) (157.38, 159.32 and 155.34 mg/100ml, respectively) than the others. There was no significant effect on high density lipoproteins (HDL) concentration and on the pH values with feeding on the experimental rations.

Blood Plasma cholesterol concentration is known to be influenced by the quantity and quality of fat in the diet. Low density lipoproteins (LDL) and cholesterol concentrations fall when saturated triglycerides in the diet is replaced by polyunsaturated vegetable oil (Spady and Woollett, 1990). Triglyceride (TG) is transported in blood via macromolecular particles called lipoproteins (Kleppe et al., 1988).

It has been concern that a substantial increase in carbohydrate containing food at the expense of fat, might result in a decreased in high-density lipoprotein and a corresponding increase in very low-density lipoprotein and triglycerides in the blood (Hicks et al., 1990).

Conclusively, agro-industrial by-products such as DDGS without supplemented seaweed could be increased urea-N in blood. Feeding on Distiller’s dried grains with soluble led to increase the low density lipoprotein, triglycerides, cholesterol and total lipids in blood plasma of growing rabbits.

REFERENCES


تأثير إحلال النواتج العرضية على تفتيير الحبوب الحافة بالسوائل محل شعر
الشعر وكسب قول الصويا مع إضافة أو بدون إضافة الطحالب البحرية في
علاقين الأرنب النامية على:

- الماكول وامتصاص الكالسيوم والفسفور وبعض مكونات الدم في الأرنب النامية.

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معهد بحوث الإنتاج الحيواني - مركز البحوث الزراعية - الدقي - ج.م.ع.
قسم إنتاج الحيوان - كلية الزراعة - جامعة المنصورة - المنصورة - ج.م.ع.

تم اختيار 27 نابورا نيوزيلندي أبيض عمر 7 أسابيع بمتوسط وزن 18±0.10
(جم ومتوسط عضلات في تسع مجاعين متساوية في العدد والعم ووزن الجسم 3
أربعة / مجموعة). استخدمت الأربعة لتقييم تأثير كلاكسوم والفسفور في الدم وذك ذلك عند
عمر 7، 9، 11، 13 أسبوع ثم أخذت عينات الدم عند الأسبوع 13 لتقييم مكونات الدم
المختلفة. ثم أخذ جزء من الأمعاء الدقيقة 5 سم بعد ذبح الأرناب وذلك لتقييم معدل
امتصاص كل من الكالسيوم والفسفور.

تم تغذية الأرناب على تسع علاقات تجريبية على النحو التالي: العلقة (الأولى)
المقارنة تحتوي على: 10% أنثى صفرا = 10% شعر + 13.7% كسب قول
الصويا = 20% نخالة = 4% حجر جيري + 1% كاسبيوم فوسفات + 3% جمع طعام + 4% مخلوط معنئ
وقيقينات + 2% نابورا.

وتم إجراء النواتج العرضية تستجابة الحبوب الحافة بالسوائل محل الشعر وكسب
قول الصويا بنسبة 10% (كل من النسبة مقسمة على كل من الشعر وكسب قول
الصويا بنسبة متساوية) وذلك لعلاقين (الثانية والثانية). تم تم إضافة الطحالب البحرية لهذه
العلاقين بنسبة 5% وذلك لكل من العلقة الرابعة والخامسة والسادسة. كما تم إضافة
الطحالب البحرية لهذه العلاقين بنسبة 1% وذلك لكل من العلقة السابعة والثامنة والتاسعة.
وكان جميع العلاقات على شكل مكعبات ومتساوية في الطاقة والبروتين تقريباً وطبقا
للاحتياجات المطلوبة تغذية الأرناب النامية.

- زادت كمية الماكول من الكالسيوم معونياً (0.05) عند التغذية على العلقة الثامنة
(192 جم/رس/اليوم) مقارنة بالتغذية على العلائق الأخرى. بينما زادت
كمية المأكولات من الفسفر معونيا (2005) عند التغذية على العلبة التاسعة،
الخامسة، السادسة، والثامنة (96, 94، 96 و 95 مليم/ رأس/ اليوم، على التوالي) مقارنة بالزمن على العلائق الأخرى.
• لم تظهر فروق معنوية على إنتحام الكالسيوم والفسفر أو على تركيز كل
منها في الدم نتيجة للمعالات التجريبية المختلفة.
• زاد تركيز اليوريا في الدم معونيا (2005) عند التغذية على العلبة الثانية، الثالثة
و السابعة (0.37 و 0.35 مليم/ 100 ملي، على التوالي) مقارنة بالزمن على العلائق التجريبية الأخرى.
• زاد تركيز الكوليسترول في الدم معونيا (2005) عند التغذية على العلبة التاسعة
(234 مليم/ 100 ملي) مقارنة بالزمن على العلائق التجريبية الأخرى.
• زاد تركيز الجليسيريدات الثلاثية في بلازما الدم معونيا (2005) عند التغذية على
العلبة الثانية، الخامسة، السادسة، والثامنة (26, 20 و 15 مليم/ 100 ملي، على التوالي) مقارنة بالزمن على العلائق الأخرى. كما كان تركيز الدهون الكلية مرتفعا ومعونيا (2005) عند
الزمن على نفس العلائق السابقة وكانت القيم المتحصل عليها (32 و 25 مليم/ 100 ملي، على التوالي) مقارنة بالزمن على العلائق التجريبية الأخرى.
• زاد تركيز البروتينات الدهنية منخفضة الكثافة في الدم معونيا (2005) عند
الزمن على العلبة التاسعة (123 مليم/ 100 ملي) مقارنة بالزمن على
العلاق التجريبية الأخرى.
التوصية: بسبيق يتضح أن التغذية على النواتج الوعائية لتقطير الحبوب الجافة
بالسواح بدون إضافة الطحالب البحرية أدت إلى زيادة تركيز اليبوز والجلسرادات
الثالثة والكوليسترول والدهون الكلية بالدم.